

**Brandon Mark Gassaway, PhD**  
**Assistant Professor**  
E111 BNSN, Brigham Young University  
Provo, UT 84602  
801-422-7193  
[brandon.gassaway@byu.edu](mailto:brandon.gassaway@byu.edu)

---

## Education

---

- Doctor of Philosophy in Cellular and Molecular Physiology** *May 2018*  
*Yale University, New Haven, CT*
- Thesis Accepted with Distinction
  - Certificate of Completion, Medical Research Scholars Program
- B.S. Degree in Biochemistry** *April 2011*  
*Brigham Young University, Provo, UT*
- *Magna Cum Laude* with University Honors

## Research Experience

---

- Assistant Professor** *Jan 2024-current*  
*Brigham Young University Department of Chemistry & Biochemistry*
- Research Focus: Functionalizing protein post-translational modifications using thermal stability proteomics and phospho-amino acid orthogonal translation systems
- Post-Doctoral Fellow, Dr. Marcia Haigis** *Sept 2021-Dec 2023*  
*Harvard Medical School Department of Cell Biology*
- Research Focus: Applying novel proteomics methods to the investigation of T cell activation and function
  - Techniques: Phosphorylation Integrated Thermal Shift Assay, Primary murine T cell isolation and culture
- Post-Doctoral Fellow, Dr. Steven Gygi** *July 2018-Dec 2023*  
*Harvard Medical School Department of Cell Biology*
- Research Focus: Developing experimental methods and bioinformatics analyses to functionalize proteins and protein phosphorylation
  - Techniques: TMT-based quantitative proteomics and phosphoproteomics, protein thermal stability, affinity-purification mass spectrometry
- Ph.D. Candidate, Dr. Jesse Rinehart** *Aug 2012-May 2018*  
*Yale Department of Cellular and Molecular Physiology, Systems Biology Institute*
- Project: investigating the mechanisms of hepatic insulin resistance by phosphoproteomic analysis
  - Techniques: quantitative proteomics and phosphoproteomics, systems analysis, enzyme activity, protein purification and analysis, phosphoserine protein production using orthogonal translation systems
- Research Intern, Dr. Lisa Elkin** *June 2012-Aug 2012*  
*Bristol-Myers-Squibb Pharmaceuticals*
- Project: development of mass spectrometry based assays for assessment of transporter inhibition
  - Techniques: LC-MS/MS (Agilent RapidFire<sup>®</sup> platform), small molecule lead evaluation, radioactive substrate uptake assays
- Research Assistant, Dr. Emily Bates** *June 2009-April 2011*  
*BYU Department of Chemistry and Biochemistry*
- Project: the role of inwardly rectifying potassium channels in *Drosophila* embryonic and adult development
  - Techniques: phenotypic analysis, *Drosophila* genetics, immunohistochemistry

## Teaching Experience

---

- Certificate of College Teaching Preparation/CIRTL Associate** *Spring 2018*

---

<b>Instructor</b> , Dr. Jesse Rinehart and Dr. Terrence Wu <i>Yale West Campus Analytical Core Proteomics Bootcamp 2018</i>	<i>Spring 2018</i>
<ul style="list-style-type: none"> <li>• Developed and presented curriculum for the design, execution, and data analysis of proteomics experiments</li> <li>• Designed and carried out hands-on instruction on mass spectrometer instrument maintenance</li> </ul>	
<b>Teaching Fellow 2</b> , Dr. Emile Boulpaep <i>Cellular and Molecular Physiology 560</i>	<i>Winter 2014</i>
<ul style="list-style-type: none"> <li>• Designed and presented review material</li> <li>• Graded homework and exams</li> </ul>	
<b>Teaching Fellow 2</b> , Dr. Stuart Campbell, Dr. W. Mark Saltzman <i>Cellular and Molecular Physiology 550</i>	<i>Fall 2012</i> <i>Fall 2013</i>
<ul style="list-style-type: none"> <li>• Designed and presented weekly review sessions using outside material and review activities</li> <li>• Helped write exam questions</li> <li>• Graded homework and exams</li> </ul>	

## **Awards, Honors, and Fellowships**

---

<b>Gruber Fellow</b> , Gruber Foundation	<i>Sept 2011</i>
<b>Graduate Research Fellow</b> , National Science Foundation	<i>April 2011</i>
<b>Kagel and Blessing Scholarship</b> , Blessing Family Foundation	<i>Jan 2010</i>
<ul style="list-style-type: none"> <li>• For non-dance majors in performing groups</li> </ul>	
<b>Heritage Scholarship</b> , Brigham Young University	<i>Aug 2005</i>
<ul style="list-style-type: none"> <li>• Four years, full tuition</li> </ul>	
<b>Most Valuable Student Scholarship</b> , National Elks Foundation	<i>April 2005</i>

## **Meetings and Presentations**

---

Speaker. 70 <sup>th</sup> Annual Conference of the American Society of Mass Spectrometry. Rigorous Benchmarking and Improvement of Phosphopeptide Identification and False Localization Rates using a Proteome-scale Synthetic Human Phosphoserine Library. Philadelphia, PA	<i>Nov 3, 2021</i>
Poster. Harvard Cell Biology Department Retreat 2019. Investigating Phosphorylation-dependent Protein-protein Interactions in the Bioplex Network. Falmouth, MA	<i>Oct 9, 2019</i>
Poster. Harvard Cell Biology Department Retreat 2018. Investigating Phosphorylation-dependent Protein-protein Interactions in the Bioplex Network. Falmouth, MA	<i>Oct 15, 2018</i>
Poster. Yale West Campus 10 <sup>th</sup> Anniversary Symposium. Proteomics methods applied to cultural heritage: Identifying proteins in an ancient Roman Shield. West Haven, CT	<i>Oct 26, 2017</i>
<ul style="list-style-type: none"> <li>• Awarded Best Poster</li> </ul>	
Poster. Yale Cellular and Molecular Physiology Department Annual Retreat. Pyruvate Kinase Phosphorylation is Regulated by Cell Cycle and Nutrient Status with an Unexpected Effect on Enzyme Activity. West Haven, CT	<i>Sep 22, 2017</i>
<ul style="list-style-type: none"> <li>• Awarded Best Poster</li> </ul>	
Poster. 76 <sup>th</sup> Scientific Sessions of the American Diabetes Association. PKC $\epsilon$ and CDKs Regulate Insulin Signaling in Response to a 3-day High Fat Diet. New Orleans, LA	<i>June 10-14, 2016</i>
Poster. 64 <sup>th</sup> Annual Conference of the American Society of Mass Spectrometry. PKC $\epsilon$ and CDKs Regulate Insulin Signaling in Response to a 3-day High Fat Diet. San Antonio, TX	<i>June 5-9, 2016</i>
Poster. 8 <sup>th</sup> United Nations Global Colloquium of University Presidents. Protein Analysis of Dura-Europos Shield Materials. West Haven, CT	<i>April 13, 2016</i>

- Poster. Yale Cellular and Molecular Physiology Department Annual Retreat. A Quantitative Phosphoproteomic Analysis of Acute Hepatic Insulin Resistance Reveals Novel Regulators of Insulin Signaling. West Haven, CT Oct 27, 2015  
 • Awarded Best Poster
- Speaker. HHMI Med-into-Grad Symposium 2015 Metabolism: Fueling Translational Research. A Quantitative Phosphoproteomic Analysis of Mechanisms of Insulin Resistance Reveals a PKC $\epsilon$ -Regulated Network. Boston, MA Oct 9-10, 2015
- Poster. Yale Systems Biology Institute Symposium & Ribbon Cutting Ceremony. A Quantitative Phosphoproteomic Analysis of Mechanisms of Insulin Resistance Reveals a PKC $\epsilon$ -Regulated Network. West Haven, CT April 22, 2015
- Poster. Yale Cellular and Molecular Physiology Department Annual Retreat. A Quantitative Phosphoproteomics Approach to Mechanisms of Hepatic Insulin Resistance II. Phosphoproteomic Analysis. West Haven, CT Sep 23, 2014
- Poster. MaxQuant Summer School: Computational Mass Spectrometry-Based Proteomics. Quantitative Phosphoproteomic Analysis of Acute Lipid Exposure in Rats Reveals a Direct Regulation of mTOR by AMPK. Bethesda MD. July 21-25, 2014
- Poster. Yale Cellular and Molecular Physiology Department Annual Retreat. Quantitative Phosphoproteomic Analysis of *abl-1* Knockout in *C. elegans* Embryos. West Haven, CT Oct 28, 2013
- Speaker. Mass Spectrometry in Biological Applications and New Technologies. Quantitative Phosphoproteomic Analysis of *abl-1* Knockout in *C. elegans* Embryos. West Haven, CT June 25, 2013

## Publications

---

- Gassaway, B.M.**, Li, J., Rad, R., Mintseris, J., Mohler, K., Levy, T., Aguiar, M., Beausoleil, S., Paulo, J.A., Rinehart, J., Huttlin, E.L., Gygi, S.P. (2022). A Multi-purpose, Regenerable, Proteome-scale, Human Phosphoserine Resource for Phosphoproteomics. *Nature Methods*, DOI: 10.1038/s41592-022-01638-5
- Ferguson, C. J., Urso, O., Bodrug, T., **Gassaway, B. M.**, Watson, E. R., Prabu, J. R., Lara-Gonzalez, P., Martinez-Chacin, R. C., Wu, D. Y., et al. (2022). APC7 mediates ubiquitin signaling in constitutive heterochromatin in the developing mammalian brain. *Molecular Cell*, 82(1), 90-105.e13.
- Huttlin, E.L., Bruckner, R.J., Navarrete-Perea, J., Cannon, J.R., Baltier, K., Gebreab, F., Gygi, M.P., Thornock, A., Zarraga, G., Tam, S., Szpyt, J., **Gassaway, B.M.**, et al. (2021). Dual proteome-scale networks reveal cell-specific remodeling of the human interactome. *Cell* 184, 3022-3040.e28.
- Apostolidi, M., Vathiotis, I.A., Muthusamy, V., Gaule, P., **Gassaway, B.M.**, Rimm, D.L., and Rinehart, J. (2021). Targeting Pyruvate Kinase M2 Phosphorylation Reverses Aggressive Cancer Phenotypes. *Cancer Research* 81, 4346–4359.
- Gassaway, B.M.**, Paulo, J.A., and Gygi, S.P. (2021). Categorization of Phosphorylation Site Behavior during the Diauxic Shift in *Saccharomyces cerevisiae*. *Journal of Proteome Research* 20, 2487–2496.
- Ringel, A.E., Drijvers, J.M., Baker, G.J., Catozzi, A., Garcia-Canaveras, J.C., **Gassaway, B.M.**, Miller, B.C., Juneja, V.R., Nguyen, T.H., Joshi, S., et al. (2020). Obesity Shapes Metabolism in the Tumor Microenvironment to Suppress Anti-Tumor Immunity. *Cell* 183, 1848-1866 e26.
- Wu, J., Rowart, P., Jouret, F., **Gassaway, B.M.**, Rajendran, V., Rinehart, J., and Caplan, M.J. (2020). Mechanisms involved in AMPK-mediated deposition of tight junction components to the plasma membrane. *American Journal of Physiology-Cell Physiology* 318, C486–C501.
- Gassaway, B.M.**, Cardone, R.L., Padyana, A.K., Petersen, M.C., Judd, E.T., Hayes, S., Tong, S., Barber, K.W., Apostolidi, M., Abulizi, A., et al. (2019). Distinct Hepatic PKA and CDK Signaling Pathways Control Activity-Independent Pyruvate Kinase Phosphorylation and Hepatic Glucose Production. *Cell Rep* 29, 3394-3404.e9.
- Codina, A., Renauer, P.A., Wang, G., Chow, R.D., Park, J.J., Ye, H., Zhang, K., Dong, M.B., **Gassaway, B.**, Ye, L., et al. (2019). Convergent Identification and Interrogation of Tumor-Intrinsic Factors that Modulate Cancer Immunity In Vivo. *Cell Syst* 8, 136-151 e7.

- Samuel, V.T., Petersen, M.C., **Gassaway, B.M.**, Vatner, D.F., Rinehart, J., and Shulman, G.I. (2019). Considering the Links Between Nonalcoholic Fatty Liver Disease and Insulin Resistance: Revisiting the Role of Protein Kinase C  $\epsilon$ . *Hepatology* 70, 2217–2220.
- Gassaway, B.M.**, Petersen, M.C., Surovtseva, Y. v, Barber, K.W., Sheetz, J.B., Aerni, H.R., Merkel, J.S., Samuel, V.T., Shulman, G.I., and Rinehart, J. (2018). PKCepsilon contributes to lipid-induced insulin resistance through cross talk with p70S6K and through previously unknown regulators of insulin signaling. *Proc Natl Acad Sci U S A* 115, E8996–E9005.
- D’Lima, N.G., Khitun, A., Rosenbloom, A.D., Yuan, P., **Gassaway, B.M.**, Barber, K.W., Rinehart, J., and Slavoff, S.A. (2017). Comparative Proteomics Enables Identification of Nonannotated Cold Shock Proteins in *E. coli*. *J Proteome Res* 16, 3722–3731.
- Mohler, K., Aerni, H.R., **Gassaway, B.**, Ling, J., Ibba, M., and Rinehart, J. (2017a). MS-READ: Quantitative measurement of amino acid incorporation. *Biochim Biophys Acta Gen Subj* 1861, 3081–3088.
- Mohler, K., Mann, R., Bullwinkle, T.J., Hopkins, K., Hwang, L., Reynolds, N.M., **Gassaway, B.**, Aerni, H.R., Rinehart, J., Polymenis, M., et al. (2017b). Editing of misaminoacylated tRNA controls the sensitivity of amino acid stress responses in *Saccharomyces cerevisiae*. *Nucleic Acids Res* 45, 3985–3996.
- Ferdaus, M.Z., Barber, K.W., Lopez-Cayuqueo, K.I., Terker, A.S., Argaiz, E.R., **Gassaway, B.M.**, Chambrey, R., Gamba, G., Rinehart, J., and McCormick, J.A. (2016). SPAK and OSR1 play essential roles in potassium homeostasis through actions on the distal convoluted tubule. *J Physiol* 594, 4945–4966.
- Petersen, M.C., Madiraju, A.K., **Gassaway, B.M.**, Marcel, M., Nasiri, A.R., Butrico, G., Marcucci, M.J., Zhang, D., Abulizi, A., Zhang, X.M., et al. (2016). Insulin receptor Thr1160 phosphorylation mediates lipid-induced hepatic insulin resistance. *J Clin Invest* 126, 4361–4371.
- Rovner, A.J., Haimovich, A.D., Katz, S.R., Li, Z., Grome, M.W., **Gassaway, B.M.**, Amiram, M., Patel, J.R., Gallagher, R.R., Rinehart, J., et al. (2015). Recoded organisms engineered to depend on synthetic amino acids. *Nature* 518, 89–93.
- Sawyer, N., **Gassaway, B.M.**, Haimovich, A.D., Isaacs, F.J., Rinehart, J., and Regan, L. (2014). Designed phosphoprotein recognition in *Escherichia coli*. *ACS Chem Biol* 9, 2502–2507.
- Loffler, M.G., Birkenfeld, A.L., Philbrick, K.M., Belman, J.P., Habtemichael, E.N., Booth, C.J., Castorena, C.M., Choi, C.S., Jornayvaz, F.R., **Gassaway, B.M.**, et al. (2013). Enhanced fasting glucose turnover in mice with disrupted action of TUG protein in skeletal muscle. *J Biol Chem* 288, 20135–20150.
- Dahal, G.R., Rawson, J., **Gassaway, B.**, Kwok, B., Tong, Y., Ptacek, L.J., and Bates, E. (2012). An inwardly rectifying K<sup>+</sup> channel is required for patterning. *Development* 139, 3653–3664.

## Personal Information

---

- Member, American Society of Mass Spectrometry
- Scoutmaster, Boy Scouts of America 2012-2015
- Performer, BYU International Folk Dance Ensemble 2009-2011
- Eagle Scout 2001
- Conversational Proficiency in Ukrainian and Russian